

Enterococci qPCR: Potential Impacts to POTWs and Permit Compliance

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POTW Perspectives

- Water Quality Standards to Permit Process
- Implementation
- Practical Reality

Water Quality Standards to NPDES

- The process begins when a state or Indian tribe establishes water quality standards for a water body within its jurisdiction, as required by the Clean Water Act.
 - Designated Use
 - Water Quality Criterion
 - Policy of preventing degradation of the water body

Water Quality Standards to NPDES

- NPDES effluent limits are established to insure that the point source discharges of pollutants treat to a level that do not cause the waterbody to exceed the established water quality standard.

Water Quality Standards to NPDES

- Effluent monitoring allows the NPDES authority to assess compliance with the required controls.
- Next, there is some process to assess the waterbody to determine if it is attaining the established standards.

Water Quality Standards to NPDES

- The point of this monitoring is to evaluate the condition of the water body and to establish a cause-and-effect relationship.
- In order to establish a cause-and-effect relationship there must be congruency between the cause and the effect.

Water Quality Standards to NPDES

- In order to establish a cause-and-effect relationship the interpretation the genome equivalents for Enterococci must be accurately converted to the number of colony forming units.

Water Quality Standards to NPDES

- Any proposed criterion must measure the efficacy of the treatment process.
- The results of a WERF Final Report entitled, "Molecular Alternatives to Indicator and Pathogen Detection: Real-Time PCR", indicated that the real time result is inadequate to represent the efficacy of sewage treatment.

Table 9-2. Comparison of *Enterococcus* Quantification by Real-time PCR and Culture Assay in Environmental Waters.

| Sample | Real-time PCR (Genomic copies/100 ml) | Culture assay* (CFU or MPN/100 ml) |
|---|--|---------------------------------------|
| Sewage | | |
| 9/15/03 | | |
| Primary effluent–Plant I | 3.5x10 ⁵ | 8.1x10 ⁵ |
| Secondary effluent–Plant I | 1.4x10 ⁵ | 6.0x10 ⁵ |
| Secondary effluent–Plant II | 1.7x10 ⁵ | 3.2x10 ⁴ |
| 9/30/03 | | |
| Primary effluent–Plant I | 2.6x10 ⁶ | 1.8x10 ⁶ |
| Secondary effluent–Plant I | 3.5x10 ⁵ | 1.4x10 ⁵ |
| Primary effluent–Plant II | 1.6x10 ⁵ | 1.7x10 ⁵ |
| Secondary effluent with chlorine–Plant II | 6.9x10 ⁵ | 0 |
| 10/1/03 | | |
| Primary effluent–Plant I | 6.4x10 ⁵ | 2.0x10 ⁶ |
| Secondary effluent–Plant I | 2.1x10 ⁵ | 2.3x10 ⁴ |
| Primary effluent–Plant II | 8.1x10 ⁵ | 4.2x10 ⁵ |
| Secondary effluent with chlorine–Plant II | 5.4x10 ⁵ | 0 |
| Coastal Waters | | |
| San Diego Creek | 4.5x10 ³ | 8.8x10 ² |
| Mid Newport Bay | 2.4x10 ³ | 7.2 |
| Lower Newport Bay | 7.3x10 ² | 59 |
| Newport Dunes (multiple sites n=4) | (2.8±2.0) x 10 ² | Below detection** |
| Newport Dunes (multiple sites n=27) | Below detection | Below detection |

*All samples except Newport Dunes samples were determined by membrane filtration; Newport Dunes samples were determined using Idexx kit.

**Detection limit for real-time PCR is 5 CFU/reaction, for culturing assay is 10 MPN/100 ml

Implementation

- How can we establish a relationship between a source and any water quality impairment?
- What non-attainment of standards will cause an impairment?
- How will waste load allocations be established?

Implementation

- How do we compare the results of culture-based methods and the qPCR method?
- How does the levels of protection compare?
- How do we justify the “need” for a rapid method for wastewater effluents with the cost of implementation.

Implementation

- Training & Resources
- Public Acceptance

Practical Reality



Instructions on Bic Pipet

1. Remove back plug of pen
2. Grab ink head with pliers and gently remove internal ink tube from pen
(you now have a Bic pipet ready for use)
3. Stick bic pipet into solids and press forefinger at back of pen to create pipeting action
4. Allow one drop from bic pipet on microscope slide and be sure no ink is in sample
5. Cover drop
6. View under microscope

